

# Liberación y metabolismo del ácido palmitoleico y sus isómeros posicionales en macrófagos de ratón

Jesús Balsinde, et al.

*Instituto de Biología y Genética Molecular, Consejo Superior de Investigaciones Científicas (CSIC),  
47003 Valladolid, Spain, and*

*Centro de Investigación Biomédica en Red de Diabetes y Enfermedades Metabólicas Asociadas (CIBERDEM),  
28029 Madrid, Spain*

October 15, 2020

Los isómeros posicionales del ácido hexadecenoico han demostrado poseer propiedades antiinflamatorias. El más conocido de ellos, el ácido palmitoleico (ácido cis-9-hexadecenoico, 16:1n-7), ha sido identificado como una lipoquina con importantes acciones beneficiosas en enfermedades metabólicas, y el ácido hipogeico (ácido cis-7-hexadecenoico, 16:1n-9) puede ser considerado como un posible biomarcador de la formación de células espumosas durante la aterosclerosis. A pesar de la importancia de estos isómeros como posibles reguladores de las respuestas inflamatorias, se sabe muy poco sobre la regulación de sus niveles y distribución y movilización entre los diferentes grupos de lípidos dentro de la célula. En este trabajo se describe que la mayor parte de los ácidos grasos hexadecenoicos que se encuentran en los macrófagos peritoneales de ratón se esterifican en una especie única de fosfatidilcolina, que contiene ácido palmítico en la posición sn-1, y ácido hexadecenoico en la posición sn-2. Esta especie disminuye notablemente cuando los macrófagos se activan con estímulos inflamatorios, en paralelo con la movilización neta de ácido hexadecenoico libre. Utilizando inhibidores farmacológicos y silenciamiento génico, demostramos que los ácidos hexadecenoicos son liberados selectivamente por la fosfolipasa A<sub>2</sub> independiente de Ca<sup>2+</sup> de grupo VIA en condiciones de activación. Mientras que la mayoría del ácido hexadecenoico liberado se acumula en forma libre, una parte significativa también se transfiere a otros fosfolípidos para formar fosfolípidos de inositol que contienen hexadecenoato. También se utiliza para formar ésteres de ácidos grasos con hidroxiácidos grasos, compuestos con propiedades antidiabéticas y antiinflamatorias. Colectivamente, estos datos revelan nuevas vías y mecanismos para la utilización de ácido palmitoleico y sus isómeros bajo condiciones inflamatorias, y sugieren que parte de la actividad antiinflamatoria de dichos ácidos grasos puede deberse a su conversión en otros mediadores lipídicos.

Financiación: Ministerio de Economía, Industria y Competitividad (SAF2016-80883-R)  
Ministerio de Ciencia e Innovación (PID2019-105989RB-I00)

## REFERENCES

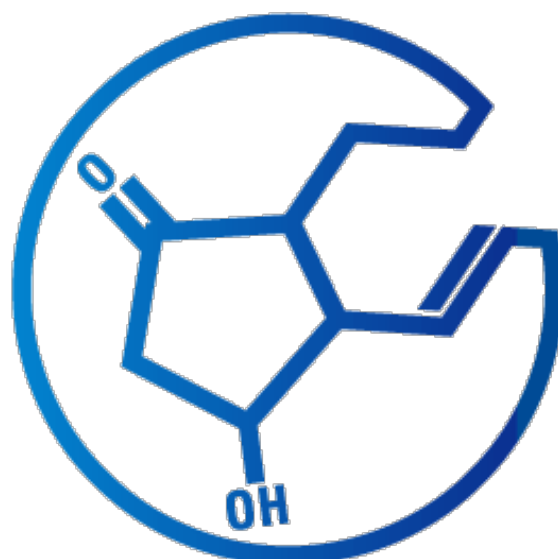
1. Guijas, C., G. Pérez-Chacón, A. M. Astudillo, J. M. Rubio, L. Gil-de-Gómez, M. A. Balboa, and J. Balsinde. 2012. Simultaneous activation of p38 and JNK by arachidonic acid stimulates the cytosolic phospholipase A<sub>2</sub>-dependent synthesis of lipid droplets in human monocytes. *J. Lipid Res.* 53: 2343–2354.
2. Guijas, C., C. Meana, A. M. Astudillo, M. A. Balboa, and J. Balsinde. 2016. Foamy monocytes are enriched in cis-7-hexadecenoic fatty acid (16:1n-9), a possible biomarker for early detection of cardiovascular disease. *Cell Chem. Biol.* 23: 689–699.
3. Astudillo, A. M., C. Meana, C. Guijas, L. Pereira, R. Lebrero, M. A. Balboa, and J. Balsinde. 2018.

- Occurrence and biological activity of palmitoleic acid isomers in phagocytic cells. *J. Lipid Res.* 59: 237–249.
4. Guijas, C., M.A. Bermúdez, C. Meana, A.M. Astudillo, L. Pereira, L. Fernández-Caballero, M.A. Balboa, and J. Balsinde. 2019. Neutral lipids are not a source of arachidonic acid for lipid mediator signaling in human foamy monocytes. *Cells* 8: 941.
  5. Pérez-Chacón, G., A. M. Astudillo, D. Balgoma, M. A. Balboa, and J. Balsinde. 2009. Control of free arachidonic acid levels by phospholipases A<sub>2</sub> and lysophospholipid acyltransferases. *Biochim. Biophys. Acta* 1791: 1103–1113.
  6. Astudillo, A. M., D. Balgoma, M. A. Balboa, and J. Balsinde. 2012. Dynamics of arachidonic acid mobilization by inflammatory cells. *Biochim. Biophys. Acta* 1821: 249–256.
  7. Guijas, C., J. P. Rodríguez, J. M. Rubio, M. A. Balboa, and J. Balsinde. 2014. Phospholipase A<sub>2</sub> regulation of lipid droplet formation. *Biochim. Biophys. Acta* 1841: 1661–1671.
  8. Astudillo, A. M., M. A. Balboa, and J. Balsinde. 2019. Selectivity of phospholipid hydrolysis by phospholipase A<sub>2</sub> enzymes in activated cells leading to polyunsaturated fatty acid mobilization. *Biochim. Biophys. Acta* 1864: 772–783.
  9. Gil-de-Gómez, L., A. M. Astudillo, C. Guijas, V. Magrioti, G. Kokotos, M. A. Balboa, and J. Balsinde. 2014. Cytosolic group IVA and calcium-independent group VIA phospholipase A<sub>2</sub>s act on distinct phospholipid pools in zymosan-stimulated mouse peritoneal macrophages. *J. Immunol.* 192: 752–762.
  10. Monge, P., A. Garrido, J.M. Rubio, V. Magrioti, G. Kokotos, M.A. Balboa, and J. Balsinde, J. 2020. The contribution of cytosolic group IVA and calcium-independent group VIA phospholipase A<sub>2</sub>s to adrenergic acid mobilization in murine macrophages. *Biomolecules* 10: 542.
  11. Pérez, R., X. Matabosch, A. Llebaria, M.A. Balboa, and J. Balsinde. 2006. Blockade of arachidonic acid incorporation into phospholipids induces apoptosis in U937 promonocytic cells. *J. Lipid Res.* 47: 484–491.
  12. Balsinde, J., B. Fernández, and E. Diez. 1990. Regulation of arachidonic acid release in mouse peritoneal macrophages. The role of extracellular calcium and protein kinase C. *J. Immunol.* 144: 4298–4304.
  13. Pindado, J., J. Balsinde, and M. A. Balboa. 2007. TLR3-dependent induction of nitric oxide synthase in RAW 264.7 macrophage-like cells via a cytosolic phospholipase 2/cyclooxygenase-2 pathway. *J. Immunol.* 179: 4821–4828.
  14. Ruipérez, V., A. M. Astudillo, M. A. Balboa, and J. Balsinde. 2009. Coordinate regulation of TLR-mediated arachidonic acid mobilization in macrophages by group IVA and group V phospholipase A<sub>2</sub>s. *J. Immunol.* 182: 3877–3883.
  15. Balsinde, J., B. Fernández, J.A. Solís-Herruzo, and E. Diez. 1992. Pathways for arachidonic acid mobilization in zymosan-stimulated mouse peritoneal macrophages. *Biochim. Biophys. Acta* 1136: 75–82.
  16. Balboa, M. A., R. Pérez, and J. Balsinde. 2003. Amplification mechanisms of inflammation: paracrine stimulation of arachidonic acid mobilization by secreted phospholipase A<sub>2</sub> is regulated by cytosolic phospholipase A<sub>2</sub>-derived hydroperoxyeicosatetraenoic acid. *J. Immunol.* 171: 989–994.
  17. Balboa, M.A., R. Pérez, and J. Balsinde. 2008. Calcium-independent phospholipase A<sub>2</sub> mediates proliferation of human promonocytic U937 cells. *FEBS J.* 275: 1915–1924.
  18. Balsinde, J., M.A. Balboa, P.A. Insel, and E.A. Dennis. 1997. Differential regulation of phospholipase D and phospholipase A<sub>2</sub> by protein kinase C in P388D<sub>1</sub> macrophages. *Biochem. J.* 321: 805–809.
  19. Diez, E., J. Balsinde, M. Aracil, and A. Schüller. 1987. Ethanol induces release of arachidonic acid but not synthesis of eicosanoids in mouse peritoneal macrophages. *Biochim. Biophys. Acta* 921: 82–89.
  20. Astudillo, A. M., G. Pérez-Chacón, D. Balgoma, L. Gil-de-Gómez, V. Ruipérez, C. Guijas, M. A. Balboa, and J. Balsinde. 2011. Influence of cellular arachidonic acid levels on phospholipid remodeling and CoA-independent transacylase activity in human monocytes and U937 cells. *Biochim. Biophys. Acta* 1811: 97–103.
  21. Valdearcos, M., E. Esquinas, C. Meana, L. Gil-de-Gómez, C. Guijas, J. Balsinde, and M. A. Balboa. 2011.

- Subcellular localization and role of lipin-1 in human macrophages. *J. Immunol.* 186: 6004–6013.
22. Rodríguez, J. P., C. Guijas, A. M. Astudillo, J. M. Rubio, M. A. Balboa, and J. Balsinde. 2019. Sequestration of 9-hydroxystearic acid in FAHFA (fatty acid esters of hydroxy fatty acids) as a protective mechanism for colon carcinoma cells to avoid apoptotic cell death. *Cancers* 11: 524.
  23. Balsinde, J., M.A. Balboa, and E.A. Dennis. 1997. Antisense inhibition of group VI Ca<sub>2+</sub>-independent phospholipase A<sub>2</sub> blocks phospholipid fatty acid remodeling in murine P388D<sub>1</sub> macrophages. *J. Biol. Chem.* 272: 29317–29321.
  24. Balsinde, J., M.A. Balboa, and E.A. Dennis. 2000. Identification of a third pathway for arachidonic acid mobilization and prostaglandin production in activated P388D<sub>1</sub> macrophage-like cells. *J. Biol. Chem.* 275: 22544–22549.
  25. Balboa, M. A., Y. Sáez, and J. Balsinde. 2003. Calcium-independent phospholipase A<sub>2</sub> is required for lysozyme secretion in U937 promonocytes. *J. Immunol.* 170: 5276–5280.
  26. Pérez, R., R. Melero, M.A. Balboa, and J. Balsinde. 2004. Role of group VIA calcium-independent phospholipase A<sub>2</sub> in arachidonic acid release, phospholipid fatty acid incorporation, and apoptosis in U937 cells responding to hydrogen peroxide. *J. Biol. Chem.* 279: 40385–40391.
  27. Balboa, M. A., J. Balsinde, S. S. Jones, and E. A. Dennis. 1997. Identity between the Ca<sup>2+</sup>-independent phospholipase A<sub>2</sub> enzymes from P388D<sub>1</sub> macrophages and Chinese hamster ovary cells. *J. Biol. Chem.* 272: 8576–8580.
  28. Balgoma, D., A. M. Astudillo, G. Pérez-Chacón, O. Montero, M. A. Balboa, and J. Balsinde. 2010. Markers of monocyte activation revealed by lipidomic profiling of arachidonic acid-containing phospholipids. *J. Immunol.* 184: 3857–3865.
  29. Rubio, J. M., J. P. Rodríguez, L. Gil-de-Gómez, C. Guijas, M. A. Balboa, and J. Balsinde. 2015. Group V secreted phospholipase A<sub>2</sub> is up-regulated by interleukin-4 in human macrophages and mediates phagocytosis via hydrolysis of ethanolamine phospholipids. *J. Immunol.* 194: 3327–3339.
  30. Balsinde, J., and M.A. Balboa. 2005. Cellular regulation and proposed biological functions of group VIA calcium-independent phospholipase A<sub>2</sub> in activated cells. *Cell. Signal.* 17: 1052–1062.
  31. Astudillo, A. M., G. Pérez-Chacón, C. Meana, D. Balgoma, A. Pol, M. A. del Pozo, M. A. Balboa, and J. Balsinde. 2011. Altered arachidonate distribution in macrophages from caveolin-1 null mice leading to reduced eicosanoid synthesis. *J. Biol. Chem.* 286: 35299–35307.
  32. Lebrero, P., A.M. Astudillo, J.M. Rubio, L. Fernández-Caballero, G. Kokotos, M.A. Balboa, and J. Balsinde. 2019. Cellular plasmalogen content does not influence arachidonic acid levels or distribution in macrophages: a role for cytosolic phospholipase A<sub>2</sub>γ in phospholipid remodeling. *Cells* 8: 799.
  33. Rodríguez, J.P., E. Leiguez, C. Guijas, B. Lomonte, J.M. Gutiérrez, C. Teixeira, M.A. Balboa, and J. Balsinde. 2020. A lipidomic perspective of the action of group IIA secreted phospholipase A<sub>2</sub> on human monocytes: lipid droplet biogenesis and activation of cytosolic phospholipase A<sub>2</sub>α. *Biomolecules* 10: 891.
  34. Gil-de-Gómez, L., P. Monge, J.P. Rodríguez, A.M. Astudillo, M.A. Balboa, and J. Balsinde. 2020. Phospholipid arachidonic acid remodeling during phagocytosis in mouse peritoneal macrophages. *Biomedicines* 8: 274.
  35. Gil-de-Gómez, L., A. M. Astudillo, C. Meana, J. M. Rubio, C. Guijas, M. A. Balboa, and J. Balsinde. 2013. A phosphatidylinositol species acutely generated by activated macrophages regulates innate immune responses. *J. Immunol.* 190: 5169–5177.
  36. Gil-de-Gómez, L., A. M. Astudillo, P. Lebrero, M. A. Balboa, and J. Balsinde. 2017. Essential role for ethanolamine plasmalogen hydrolysis in bacterial lipopolysaccharide priming of macrophages for enhanced arachidonic acid release. *Front. Immunol.* 8: 1251.
  37. Rubio, J. M., A. M. Astudillo, J. Casas, M. A. Balboa, and J. Balsinde. 2018. Regulation of phagocytosis in macrophages by membrane ethanolamine plasmalogens. *Front. Immunol.* 9: 1723.
  38. Balboa, M. A., J. Balsinde, E. A. Dennis, and P. A. Insel. 1995. A phospholipase D-mediated pathway for generating diacylglycerol in nuclei from Madin-Darby canine kidney cells. *J. Biol. Chem.* 270: 11738–

11740.

39. Balsinde, J., and E. A. Dennis. 1996. The incorporation of arachidonic acid into triacylglycerol in P388D<sub>1</sub> macrophage-like cells. *Eur. J. Biochem.* 235: 480–485.
40. Gubern, A., M. Barceló, D. Barneda, J. M. López, R. Masgrau, F. Picatoste, C. E. Chalfant, J. Balsinde, M. A. Balboa, and E. Claro. 2009. JNK and ceramide kinase govern the biogenesis of lipid droplets through activation of group IVA phospholipase A<sub>2</sub>. *J. Biol. Chem.* 284: 32359–32369.
41. Balsinde, J. 2002. Roles of various phospholipases A<sub>2</sub> in providing lysophospholipid acceptors for fatty acid phospholipid incorporation and remodelling. *Biochem. J.* 364: 695–702.
42. Balboa, M. A., and J. Balsinde. 2002. Involvement of calcium-independent phospholipase A<sub>2</sub> in hydrogen peroxide-induced accumulation of free fatty acids in human U937 cells. *J. Biol. Chem.* 277: 40384–40389.
43. Balgoma, D., O. Montero, M. A. Balboa, and J. Balsinde. 2010. Lipidomic approaches to the study of phospholipase A<sub>2</sub>-regulated phospholipid fatty acid incorporation and remodeling. *Biochimie* 92: 645–650.



**THE EICOSANOID  
RESEARCH DIVISION**  
V A L L A D O L I D