

Estudios sobre el origen del ácido araquidónico liberado por monocitos espumosos

Jesús Balsinde, et al.

*Instituto de Biología y Genética Molecular, Consejo Superior de Investigaciones Científicas (CSIC),
47003 Valladolid, Spain, and*

*Centro de Investigación Biomédica en Red de Diabetes y Enfermedades Metabólicas Asociadas (CIBERDEM),
28029 Madrid, Spain*

August 20, 2019

Los monocitos humanos expuestos a ácido araquidónico libre (AA), un producto liberado por las células endoteliales, adquieren un fenotipo espumoso que se debe a la acumulación de gotas lipídicas ricas en AA en el citoplasma. En este trabajo se ha investigado el posible papel del AA almacenado en los lípidos neutros de los monocitos espumosos para ser liberados por lipasas y contribuir de este modo a la señalización mediada por lípidos. Para ello, se han utilizado enfoques lipidómicos basados en espectrometría de masas combinados con estrategias para generar monocitos con diferentes concentraciones celulares de AA. Los resultados de los experimentos indican que la cantidad de AA en fosfolípidos en monocitos es relativamente estable y no cambia en gran medida al exponer las células al AA externo. Por el contrario, los niveles de AA en triacilglicerol pueden variar notablemente, pudiendo acomodarse en esta especie lipídica una cantidad relativamente grande de ácido grasos. La estimulación de las células con zimosán opsonizado provoca la pérdida de cantidades significativas de AA. En todas las condiciones las disminuciones de AA observadas se debieron a pérdidas en fosfolípidos. No se detectó bajo ninguna condición una pérdida significativa de AA de los triacilglicérols. En experimentos usando inhibidores selectivos de la hidrólisis de fosfolípidos o triacilglicerol se confirmó que los fosfolípidos de membrana constituyen el único reservorio celular desde el que se libera AA en condiciones de estimulación celular. Por tanto, el AA presente en triacilglicérol no supone una fuente de AA para la generación de eicosanoides sino que probablemente ejerce otras funciones celulares, desconocidas en este momento.

Financiación: Ministerio de Economía, Industria y Competitividad (SAF2016-80883-R)

REFERENCES

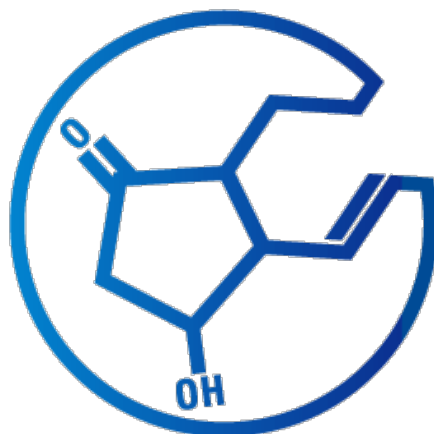
1. Guijas, C., J. P. Rodríguez, J. M. Rubio, M. A. Balboa, and J. Balsinde. 2014. Phospholipase A₂ regulation of lipid droplet formation. *Biochim. Biophys. Acta* 1841: 1661–1671.
2. Guijas, C., G. Pérez-Chacón, A. M. Astudillo, J. M. Rubio, L. Gil-de-Gómez, M. A. Balboa, and J. Balsinde. 2012. Simultaneous activation of p38 and JNK by arachidonic acid stimulates the cytosolic phospholipase A₂-dependent synthesis of lipid droplets in human monocytes. *J. Lipid Res.* 53: 2343–2354.
3. Guijas, C., C. Meana, A. M. Astudillo, M. A. Balboa, and J. Balsinde. 2016. Foamy monocytes are enriched in cis-7-hexadecenoic fatty acid (16:1n-9), a possible biomarker for early detection of cardiovascular disease. *Cell Chem. Biol.* 23: 689–699.
4. Astudillo, A. M., M. A. Balboa, and J. Balsinde. 2019. Selectivity of phospholipid hydrolysis by phospholipase A₂ enzymes in activated cells leading to polyunsaturated fatty acid mobilization. *Biochim. Biophys. Acta* 1864: 772–783.

5. Astudillo, A. M., D. Balgoma, M. A. Balboa, and J. Balsinde. 2012. Dynamics of arachidonic acid mobilization by inflammatory cells. *Biochim. Biophys. Acta* 1821: 249–256.
6. Pérez-Chacón, G., A. M. Astudillo, D. Balgoma, M. A. Balboa, and J. Balsinde. 2009. Control of free arachidonic acid levels by phospholipases A₂ and lysophospholipid acyltransferases. *Biochim. Biophys. Acta* 1791: 1103–1113.
7. Pérez-Chacón, G., A. M. Astudillo, V. Ruipérez, M. A. Balboa, and J. Balsinde. 2010. Signaling role for lysophosphatidylcholine acyltransferase 3 in receptor-regulated arachidonic acid reacylation reactions in human monocytes. *J. Immunol.* 184: 1071–1078.
8. Casas, J., C. Meana, E. Esquinas, M. Valdearcos, J. Pindado, J. Balsinde, and M. A. Balboa. 2009. Requirement of JNK-mediated phosphorylation for translocation of group IVA phospholipase A₂ to phagosomes in human macrophages. *J. Immunol.* 183: 2767–2774.
9. Balsinde, J., B. Fernández, and E. Diez. 1990. Regulation of arachidonic acid release in mouse peritoneal macrophages. The role of extracellular calcium and protein kinase C. *J. Immunol.* 144: 4298–4304.
10. Balsinde, J., B. Fernández, J.A. Solís-Herruzo, and E. Diez. 1992. Pathways for arachidonic acid mobilization in zymosan-stimulated mouse peritoneal macrophages. *Biochim. Biophys. Acta* 1136: 75–82.
11. Balsinde, J., M.A. Balboa, and E.A. Dennis. 2000. Identification of a third pathway for arachidonic acid mobilization and prostaglandin production in activated P388D₁ macrophage-like cells. *J. Biol. Chem.* 275: 22544–22549.
12. Balboa, M. A., Y. Sáez, and J. Balsinde. 2003. Calcium-independent phospholipase A₂ is required for lysozyme secretion in U937 promonocytes. *J. Immunol.* 170: 5276–5280.
13. Balsinde, J., M.A. Balboa, P.A. Insel, and E.A. Dennis. 1997. Differential regulation of phospholipase D and phospholipase A₂ by protein kinase C in P388D₁ macrophages. *Biochem. J.* 321: 805–809.
14. Balboa, M. A., R. Pérez, and J. Balsinde. 2003. Amplification mechanisms of inflammation: paracrine stimulation of arachidonic acid mobilization by secreted phospholipase A₂ is regulated by cytosolic phospholipase A₂-derived hydroperoxyeicosatetraenoic acid. *J. Immunol.* 171: 989–994.
15. Ruipérez, V., A. M. Astudillo, M. A. Balboa, and J. Balsinde. 2009. Coordinate regulation of TLR-mediated arachidonic acid mobilization in macrophages by group IVA and group V phospholipase A₂s. *J. Immunol.* 182: 3877–3883.
16. Diez, E., J. Balsinde, M. Aracil, and A. Schüller. 1987. Ethanol induces release of arachidonic acid but not synthesis of eicosanoids in mouse peritoneal macrophages. *Biochim. Biophys. Acta* 921: 82–89.
17. Astudillo, A. M., G. Pérez-Chacón, D. Balgoma, L. Gil-de-Gómez, V. Ruipérez, C. Guijas, M. A. Balboa, and J. Balsinde. 2011. Influence of cellular arachidonic acid levels on phospholipid remodeling and CoA-independent transacylase activity in human monocytes and U937 cells. *Biochim. Biophys. Acta* 1811: 97–103.
18. Astudillo, A. M., C. Meana, C. Guijas, L. Pereira, R. Lebrero, M. A. Balboa, and J. Balsinde. 2018. Occurrence and biological activity of palmitoleic acid isomers in phagocytic cells. *J. Lipid Res.* 59: 237–249.
19. Balgoma, D., A. M. Astudillo, G. Pérez-Chacón, O. Montero, M. A. Balboa, and J. Balsinde. 2010. Markers of monocyte activation revealed by lipidomic profiling of arachidonic acid-containing phospholipids. *J. Immunol.* 184: 3857–3865.
20. Gil-de-Gómez, L., A. M. Astudillo, C. Meana, J. M. Rubio, C. Guijas, M. A. Balboa, and J. Balsinde. 2013. A phosphatidylinositol species acutely generated by activated macrophages regulates innate immune responses. *J. Immunol.* 190: 5169–5177.
21. Gil-de-Gómez, L., A. M. Astudillo, C. Guijas, V. Magrioti, G. Kokotos, M. A. Balboa, and J. Balsinde. 2014. Cytosolic group IVA and calcium-independent group VIA phospholipase A₂s act on distinct phospholipid pools in zymosan-stimulated mouse peritoneal macrophages. *J. Immunol.* 192: 752–762.
22. Astudillo, A. M., G. Pérez-Chacón, C. Meana, D. Balgoma, A. Pol, M. A. del Pozo, M. A. Balboa, and J. Balsinde. 2011. Altered arachidonate distribution in macrophages from caveolin-1 null mice leading to

reduced eicosanoid synthesis. *J. Biol. Chem.* 286: 35299–35307.

23. Valdearcos, M., E. Esquinas, C. Meana, L. Gil-de-Gómez, C. Guijas, J. Balsinde, and M. A. Balboa. 2011. Subcellular localization and role of lipin-1 in human macrophages. *J. Immunol.* 186: 6004–6013.
24. Gil-de-Gómez, L., A. M. Astudillo, P. Lebrero, M. A. Balboa, and J. Balsinde. 2017. Essential role for ethanolamine plasmalogen hydrolysis in bacterial lipopolysaccharide priming of macrophages for enhanced arachidonic acid release. *Front. Immunol.* 8: 1251.
25. Rubio, J. M., J. P. Rodríguez, L. Gil-de-Gómez, C. Guijas, M. A. Balboa, and J. Balsinde. 2015. Group V secreted phospholipase A₂ is up-regulated by interleukin-4 in human macrophages and mediates phagocytosis via hydrolysis of ethanolamine phospholipids. *J. Immunol.* 194: 3327–3339.
26. Rodríguez, J. P., C. Guijas, A. M. Astudillo, J. M. Rubio, M. A. Balboa, and J. Balsinde. 2019. Sequestration of 9-hydroxystearic acid in FAHFA (fatty acid esters of hydroxy fatty acids) as a protective mechanism for colon carcinoma cells to avoid apoptotic cell death. *Cancers* 11: 524.
27. Balsinde, J. 1993. Mechanism of arachidonic acid liberation in ethanol-treated mouse peritoneal macrophages. *Biochim. Biophys. Acta* 1169: 54–58.
28. Balboa, M. A., and J. Balsinde. 2002. Involvement of calcium-independent phospholipase A₂ in hydrogen peroxide-induced accumulation of free fatty acids in human U937 cells. *J. Biol. Chem.* 277: 40384–40389.
29. Pindado, J., J. Balsinde, and M. A. Balboa. 2007. TLR3-dependent induction of nitric oxide synthase in RAW 264.7 macrophage-like cells via a cytosolic phospholipase 2/cyclooxygenase-2 pathway. *J. Immunol.* 179: 4821–4828.
30. Casas, J., M.A. Gijón, A.G. Vigo, M.S. Crespo, J. Balsinde, and M.A. Balboa. 2006. Phosphatidylinositol 4,5-bisphosphate anchors cytosolic group IVA phospholipase A₂ to perinuclear membranes and decreases its calcium requirement for translocation in live cells. *Mol. Biol. Cell* 17: 155-162.
31. Pérez, R., X. Matabosch, A. Llebaria, M.A. Balboa, and J. Balsinde. 2006. Blockade of arachidonic acid incorporation into phospholipids induces apoptosis in U937 promonocytic cells. *J. Lipid Res.* 47: 484–491.
32. Lebrero, P., A. M. Astudillo, J. M. Rubio, L. Fernández-Caballero, G. Kokotos, M. A. Balboa, and J. Balsinde. 2019. Cellular plasmalogen content does not influence arachidonic acid levels or distribution in macrophages: a role for cytosolic phospholipase A₂γ in phospholipid remodeling. *Cells* 8: 799.
33. Balsinde, J., and E. A. Dennis. 1996. The incorporation of arachidonic acid into triacylglycerol in P388D₁ macrophage-like cells. *Eur. J. Biochem.* 235: 480–485.
34. Balsinde, J., M. V. Winstead, and E. A. Dennis. 2002. Phospholipase A₂ regulation of arachidonic acid mobilization. *FEBS Lett.* 531: 2–6.
35. Balsinde, J., M. A. Balboa, S. Yedgar, and E. A. Dennis. 2000. Group V phospholipase A₂-mediated oleic acid mobilization in lipopolysaccharide-stimulated P388D₁ macrophages. *J. Biol. Chem.* 275: 4783–4786.
36. Balboa, M. A., Y. Shirai, G. Gaietta, M. H. Ellisman, J. Balsinde, and E. A. Dennis. 2003. Localization of group V phospholipase A₂ in caveolin-enriched granules in activated P388D₁ macrophage-like cells. *J. Biol. Chem.* 278: 48059–48065.
37. Meana, C., L. Peña, G. Lordén, E. Esquinas, C. Guijas, M. Valdearcos, J. Balsinde, and M. A. Balboa. 2014. Lipin-1 integrates lipid synthesis with proinflammatory responses during TLR activation in macrophages. *J. Immunol.* 193: 4614–4622.
38. Balgoma, D., O. Montero, M. A. Balboa, and J. Balsinde. 2010. Lipidomic approaches to the study of phospholipase A₂-regulated phospholipid fatty acid incorporation and remodeling. *Biochimie* 92: 645–650.
39. Guijas, C., A. M. Astudillo, L. Gil-de-Gómez, J. M. Rubio, M. A. Balboa, and J. Balsinde. 2012. Phospholipid sources for adrenic acid mobilization in RAW 264.7 macrophages: comparison with arachidonic acid. *Biochim. Biophys. Acta* 1821: 1386–1393.

40. Rubio, J. M., A. M. Astudillo, J. Casas, M. A. Balboa, and J. Balsinde. 2018. Regulation of phagocytosis in macrophages by membrane ethanolamine plasmalogens. *Front. Immunol.* 9: 1723.
41. Balsinde, J., and M.A. Balboa. 2005. Cellular regulation and proposed biological functions of group VIA calcium-independent phospholipase A₂ in activated cells. *Cell. Signal.* 17: 1052-1062.



**THE EICOSANOID
RESEARCH DIVISION**
VALLADOLID